



Carnosine and trehalose-carnosine tune the activity and expression of endogenous protection factors and their crosstalk with metal homeostasis.

Francesco Attanasio,^a Irina Naletova,^a Valentina Greco,^b Sebastiano Sciuto,^b Enrico Rizzarelli.^{a,b}

Carnosine (β -alanyl-L-histidine) is a natural dipeptide widely distributed in mammalian tissues and presented at high concentrations (0.7–2.0mM) in the brain.¹ As reported previously, carnosine augmented the secretion and expression of various neurotrophic factors (for example, BDNF), leading to the induction of neurite growth in SY-SY5Y cells.² Moreover, carnosine glial release and neuronal utilization in CNS have been described;³ carnosine intercepts the regulatory routes of Cu homeostasis in nervous cells and tissues. Cu dysregulation imply the oxidative stress, free-radical production and functional impairment leading to neurodegeneration. Barca et al showed that the extracellular carnosine exposure influenced intracellular Cu entry and affected the key Cu-sensing system (SP1 and CTR1).⁴ On this basis, carnosine, its derivate with trehalose and potential role of copper ions were investigated in the present study. First of all, we demonstrate that trehalose-carnosine crosses the cell membrane better than carnosine and its translocation does not depend on copper ions. On the next step, we analyzed a role of carnosine and its derivative in the modulation of CREB functions in the normal and in the copper ions deprivation conditions. Previously, it has been shown that carnosine and copper alone induce CREB phosphorylation^{5,6}. Here we found that 30 min of PC12 cells incubation with trehalose-carnosine stimulates CREB phosphorylation more than carnosine alone and the level of phospho-CREB depends on the presence of copper ions in the medium. To compare the influence of trehalose-carnosine and carnosine alone on copper homeostasis, a measure of the copper transporter CTR1 and transcriptional factor SP1 expression in culture of PC12 cells was carried out.

References

- 1 A. R. Hipkiss, Carnosine, diabetes and alzheimer's disease, *Expert Review of Neurotherapeutics* 9 (5) (2009) 583–585. doi:10.1586/ern.09.32.
- 2 K. Kadooka, K. Fujii, T. Matsumoto, M. Sato, F. Morimatsu, K. Tashiro, S. Kuhara, Y. Katakura, Mechanisms and consequences of carnosine-induced activation of intestinal epithelial cells, *Journal of Functional Foods* 13 (2015) 32 – 37. doi:10.1016/j.jff.2014.12.024.
- 3 K. Bauer, Carnosine and homocarnosine, the forgotten, enigmatic peptides of the brain, *Neurochemical research* 30 (10) (2005) 1339–1345. doi:10.1007/s11064-005-8806-z.
- 4 A. Barca, S. Ippati, E. Urso, C. Vetrugno, C. Storelli, M. Maffia, A. Romano, T. Verri, Carnosine modulates the Sp1-Slc31a1/Ctr1 copper-sensing system and influences copper homeostasis in murine CNS-derived cells, *American Journal of Physiology-Cell Physiology* 316 (2) (2019) C235–C245. doi:10.1152/ajpcell.00106.2018.
- 5 K. Fujii, K. Abe, K. Kadooka, T. Matsumoto, Y. Katakura, Carnosine activates the creb pathway in caco-2 cells, *Cytotechnology* 69 (3) (2017) 523–527. doi:10.1007/Fs10616-017-0089-0.
- 6 I. Naletova, C. Satriano, A. Pietropaolo, F. Giani, G. Pandini, V. Triaca, G. Amadoro, V. Latina, P. Calissano, A. Travaglia, V. G. Nicoletti, D. La Mendola, E. Rizzarelli, The Copper(II)-Assisted Connection between NGF and BDNF by Means of Nerve Growth Factor-Mimicking Short Peptides, *Cells* 8 (4) (2019) 301. doi:10.3390/cells8040301.

^a CNR - Istituto di Cristallografia, Via Paolo Gaifami 18, 95126 Catania, Italy

^b Dipartimento di Scienze Chimiche, Università degli Studi di Catania, Viale A. Doria 6, 95125 Catania, Italy

Creative Commons Attribution - Non commerciale - Condividi allo stesso modo 4.0 Internazionale

† oral communication at 1 st Conference on Crystallography, Structural Chemistry and Biosystems, (Catania) 04-06/10/2021